

STUDY OF THE BINDING OF METAL IONS WITH PHOSPHORYLASE D FROM  
*Raphanus sativus* BY THE ESR METHOD

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In the present communication we give the results of a study of the binding of phosphorylase D from *Raphanus sativus* L. with  $\text{Ca}^{2+}$  and  $\text{Mn}^{2+}$  ions, which are activators for it and also with  $\text{Pr}^{3+}$  ions, which inhibit it [1].

To a solution of the enzyme we added increasing amounts of  $\text{Mn}^{2+}$  ions, and it was then possible to observe the ESR signal of free  $\text{Mn}^{2+}$  ions only after the concentration of Mn in the solution had reached a certain value,  $[\text{Mn}]_0$ , and then the signal rose in proportion to the concentration of  $\text{Mn}^{2+}$  ions. The initial  $[\text{Mn}]_0$  section obviously characterizes the amount of ions that have been bound to the protein and therefore were not recorded under our conditions. For the two conformers  $D_S$  and  $D_L$  obtained by separating the phosphorylase on a column with a biospecific sorbent [2] this amount was 38 and 28 ions per protein molecules, respectively.

To study the binding of  $\text{Ca}^{2+}$  ions with the enzyme, calcium was added to a solution of the enzyme saturated with  $\text{Mn}^{2+}$  ions ( $[\text{Mn}] > [\text{Mn}]_0$ ). The ESR signal for the Mn scarcely changed. When a manganese salt was added to a solution of the enzyme saturated with  $\text{Ca}^{2+}$  ions it was possible to observe a slight decrease in  $[\text{Mn}]_0$ .

On the basis of these results and statements in the literature [3], according to which  $\text{Mn}^{2+}$  ions have higher covalent characteristics than  $\text{Ca}^{2+}$  ions and, consequently, more readily form bonds with proteins, it may be concluded that there are two types of sites for the binding of this protein with metal ions: 1) sites with which both  $\text{Ca}^{2+}$  and  $\text{Mn}^{2+}$  ions can be bound, and 2) sites with which  $\text{Ca}^{2+}$  ions cannot be bound while  $\text{Mn}^{2+}$  ions can be.

Similar experiments on the binding of  $\text{Pr}^{3+}$  ions with the protein (Fig. 1) showed that  $\text{Pr}^{3+}$  ions displace  $\text{Mn}^{2+}$  ions from their complex with phosphorylase D but if, on the other hand, phosphorylase D is previously bound with  $\text{Pr}^{3+}$  ions these ions are not displaced by  $\text{Mn}^{2+}$  ions. Thus,  $\text{Pr}^{3+}$  ions inhibit phosphorylase D by blocking the binding sites of the first type. The affinity of the praseodymium ions for phosphorylase D is higher than the affinity of  $\text{Mn}^{2+}$  and  $\text{Ca}^{2+}$  ions.

The measurements were performed on a Varian ESR spectrometer at an amplitude of modulation not exceeding 3.2 G and a power fed to the resonator of not more than 20 mW at room temperature.

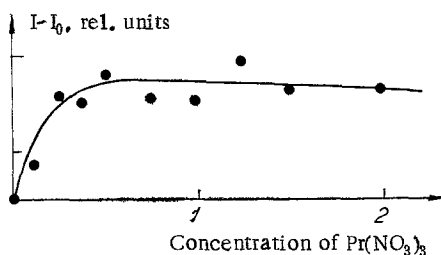


Fig. 1. Increase in the intensity of the manganese signal on the addition of a praseodymium salt to a solution of the enzyme saturated with manganese ions. Concentration of phosphorylase D 900  $\mu\text{g}$ ;  $[\text{MnCl}_2]$  0.5 mM.

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ACTIVATION OF PHOSPHORYLASE D FROM *Raphanus sativus* BY METAL IONS

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Phosphorylase D from the root-crop plant *Raphanus sativus* (garden radish) consists of an equilibrium mixture of two low-molecular-weight conformers  $D_S$  and  $D_L$  and of a high-molecular-weight form which apparently contains both the  $D_S$  and  $D_L$  conformers [1-3]. It must be mentioned that the  $D_S$  and  $D_L$  conformers differ in their catalytic properties. In the present work we have investigated the comparative influence of  $Ca^{2+}$  and  $Mn^{2+}$

The phosphorylase D was isolated from the garden radish as described previously [1]. As the substrate we used egg lecithin,  $[S] = 5$  mM, and as the initiator of the reaction sodium dodecyl sulfate (SDS), and in the determination of the transferase activity 0.4 ml of methanol was present in the medium.

As the results of the experiments performed showed,  $Ca^{2+}$  ions are more than twice as effective in transesterification (Fig. 1, curves a and d) and  $Mn^{2+}$  ions are more effective in hydrolysis (Fig. 1, curves c and d). One of the reasons why these ions exhibit different activating capacities in hydrolysis and transesterification may be the molecular heterogeneity of the phosphorylase D in solutions. It is known that  $Ca^{2+}$  ions displace the equilibrium in the direction of the formation of the high-molecular-weight variety of phosphorylase D [4, 5]. It has been shown [3] that it is precisely this form of phosphorylase D that possesses the greatest transferase activity. Furthermore, it has been shown that in the presence of  $Ca^{2+}$  ions it is just the transferase function of the enzyme that is predominant [6].

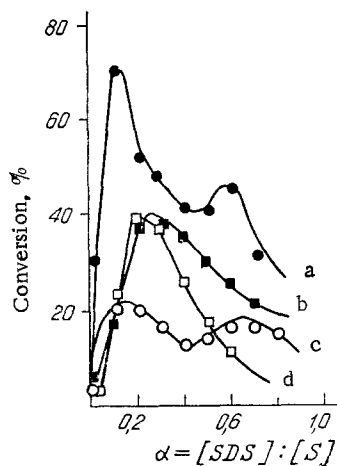


Fig. 1. Nature of the activation of phosphorylase D by  $Ca^{2+}$  ions (a, c) and  $Mn^{2+}$  ions (b, d): a, b) transesterification; c, d) hydrolysis ( $[S] = 5$  mM,  $[SDS] = 1$  mM, pH 5.6).

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